

Rat Anti-Hib-PRP IgG
(*Hemophilus influenzae* type b
Polysaccharide, PRP) ELISA Kit

Cat. No. 980-160-PRG, 96 tests

For Quantitation of Anti-Hib-PRP
IgG in Serum or plasma or other
biological fluids

For research use only, not for diagnostic or therapeutic use.



ALPHA DIAGNOSTIC
INTERNATIONAL

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ELISA Kit Components	Amount	Part
HbO-HA Coated Microwell Strip Plate	8-well strips (12)	980-131
Anti-Hib-PRP Positive Control	0.65 ml	980-132PC
Anti-Hib-PRP Calibrator 0.5 U/ml	0.65 ml	980-132A
Anti-Hib-PRP Calibrator 1 U/ml	0.65 ml	980-132B
Anti-Hib-PRP Calibrator 2.5 U/ml	0.65 ml	980-132C
Anti-Hib-PRP Calibrator 5 U/ml	0.65 ml	980-132D
Anti-Rat IgG HRP (100X)	1.5 ml	H-RtG.211
Sample Diluent (20X)	10 ml	SD20T
Low NSB Sample Diluent	30 ml	TBTm
Wash Solution Concentrate (100X)	10 ml	WB-100
TMB Substrate	12 ml	80091
Stop Solution	12 ml	80101
Product Manual	1 ea	M-980-160-PRG

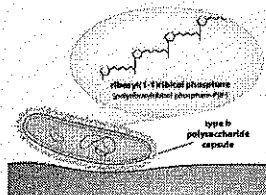
INTENDED USE

The Rat Anti-Hib-PRP IgG ELISA Kit detects and quantifies Hib-PRP (*Hemophilus influenzae* type b capsular polysaccharide) IgG in rat serum or plasma of vaccinated, immunized and/or infected hosts. This immunoassay is suitable for:

- Determining immune status relative to non-immune controls;
- Assessing efficacy of vaccines, including dosage, adjuvancy, route of immunization and timing;
- Qualifying and/or standardizing vaccine batches and protocols.

The kit has no live or killed virus or viral antigens. For research use only (RUO), not for diagnosis, cure or prevention of the disease.

GENERAL INFORMATION



Hemophilus influenzae type b (Hib) is a gram-negative coccoidal, capsular bacterium that causes, in Rats, especially in infants, bacteremia, pneumonia and acute bacterial meningitis. Vaccines prepared using the capsular polysaccharide (PRP: polyribosyl ribitol phosphate) have been effective in providing protection and minimizing the disease in populations that are routinely immunized. The PRP conjugated to various proteins, such as tetanus and diphtheria toxoid and meningococcal outer membrane protein, have been highly effective in broadening the range of immunity against disease. The Hib vaccines are also available combined with other vaccines, with a pentavalent vaccine (Hib-diphtheria-pertussis-tetanus-hepatitis B) being widely used worldwide.

The ADI Anti-Hib-PRP ELISA will quantify antibodies specific to the PRP antigen, regardless of carrier protein used for immunization.

PRINCIPLE OF THE TEST

The Rat Anti-Hib-PRP IgG ELISA kit is based on the binding of rat anti-Hib-PRP in samples to Hib-PRP immobilized on the microwells, and anti-Hib-PRP IgG antibody is detected by anti-rat IgG-specific antibody conjugated to HRP (horseradish peroxidase) enzyme. After a washing step, chromogenic substrate (TMB) is added and color is developed by the enzymatic reaction of HRP on the substrate, which is directly proportional to the amount of anti-Hib-PRP IgG present in the sample. Stopping Solution is added to terminate the reaction, and absorbance at 450nm is then measured using an ELISA microwell reader. The activity of rat antibody in samples is determined relative to anti-Hib-PRP Calibrators.

PRODUCT SPECIFICATIONS

Specificity

Purified Hib-specific polysaccharide (PRP) conjugated to human albumin (HbO-HA) is used to coat the microwells; stabilizing postcoat contains BSA; thus, no other antibody specificity is detectable in the assay. The anti-rat IgG HRP conjugate specifically detects IgG, and will not react with IgM, IgA or IgE class antibodies.

KIT CONTENTS

The microtiter well plate and all other reagents, if unopened, are stable at 2-8°C until the expiration date printed on the box label. Stabilities of the working solutions are indicated under Reagent Preparation.

To Be Reconstituted: Store as indicated.

Component	Preparation Instructions
Wash Solution Concentrate (100x) Cat. No. WB-100, 10ml	Dilute the entire volume 10ml + 990ml with distilled or deionized water into a clean stock bottle. Label as Wash Solution and store at ambient temperature until kit is used entirely.
Sample Diluent Concentrate (20x) Cat. No. SD-20T, 10ml	Dilute the entire volume, 10ml + 190ml with distilled or deionized water into a clean stock bottle. Label as Working Sample/Conjugate Diluent and store at 2-8°C until the kit lot expires or is used up.
Anti-Rat IgG - HRP Conjugate Concentrate (100x) Part No. H-RtG.211, 0.15ml	Peroxidase conjugated anti-rat IgG in buffer with protein, detergents and antimicrobial as stabilizers. Dilute fresh as needed; 10ul of concentrate to 1ml of Working Sample/Conjugate Diluent is sufficient for 1 8-well strip. Use within the working day and discard. Return 100X to 2-8°C storage.

Ready For Use: Store as indicated on labels.

Component	Part	Amt	Contents
HbO-HA Microwell Strip Plate	980-131	8-well strips (12)	Coated with recombinant Hib-PRP, and post-coated with stabilizers.
Anti-Hib-PRP Calibrators			
0.5 U/ml	980132A	0.65 ml	Four (4) vials, each containing anti-Hib-PRP in arbitrary activity Units; diluted in buffer with protein, detergents and antimicrobial as stabilizers.
1 U/ml	980132B	0.65 ml	
2.5 U/ml	980132C	0.65 ml	
5 U/ml	980132D	0.65 ml	
Anti-Hib-PRP Positive Control	980-132PC	0.65ml	Anti-HbsAg antibody; diluted in buffer with protein, detergents and antimicrobial as stabilizers. Net OD > 0.5
Low NSB Sample Diluent	TBTm Not for conjugate dilution	30 ml	Buffer with protein, detergents and antimicrobial as stabilizers. Use as is for sample dilution.
TMB Substrate	80091	12 ml	Chromogenic substrate for HRP containing TMB and peroxide.
Stop Solution	80101	12 ml	Dilute sulfuric acid.

Materials Required But Not Provided:

- Pipettors and pipettes that deliver 100ul and 1-10ml. A multi-channel pipettor is recommended.
- Disposable glass or plastic 5-15ml tubes for diluting samples and Anti-Rat IgG HRP Concentrate.
- Graduated cylinder to dilute Wash Concentrate; 200ml to 1L.
- Stock bottle to store diluted Wash Solution; 200ml to 1L.
- Distilled or deionized water to dilute reagent concentrates.
- Microwell plate reader at 450 nm wavelength.

ASSAY DESIGN AND SET-UP

Sample Collection and Handling

For serum, collect blood by venipuncture, allow clotting, and separate the serum by centrifugation at room temperature. For other samples, including tissue culture media, clarify the sample by centrifugation and/or filtration prior to dilution in Sample Diluent.

Sample Stability & Dilution

Initial dilution of serum into Working Sample Diluent (WSD) is recommended to stabilize antibody activity. This enhances reproducible sampling, and stabilizes the antibody activity for years, stored refrigerated or frozen. Further dilution into Low NSB Sample Diluent (LNSD), which provides the lowest assay background, should be at least 10 times the initial dilution and performed the same day as the assay.

Example: Initial (1/5): 10ul serum + 40ul WSD [or 0.1ml + 0.4ml]
Further (1/50): 10ul initial (1/5) + 90ul LNSD (1/50)

Assay Design

Review Calculation of Results (p5-7) and Limits of the Assay (above) before proceeding:

- Select the proper sample dilutions accounting for expected potency of positives and minimizing non-specific binding (NSB) and other matrix effects; for example, net signal for non-immune samples should be <0.5 OD. This is usually 1:50 or greater dilution for rat sera with normal levels of IgG and IgM.
- Run a Sample Diluent Blank. This signal is an indicator of proper assay performance, especially of washing efficacy, and is used for net OD calculations, if required (See Method A).
- Run the Anti-Hib-PRP Positive Control; net OD > 0.5 .
- Run a set of Calibrators. Calibrators validate that the assay was performed to specifications; results can be used to normalize between-assay variation for enhanced precision. Reading values off a Calibrator curve, Method B, has limitations. See Limits of the Assay (above).
- Run a range of sample dilutions for expected higher positives that allows calculation of antibody Titer (when specific titer is at least 4-fold higher than non-immune). See Method C.
- Run samples in duplicate if used for quantitation; non-immunes that are significantly lower than immunes may be run in singlicate. The Calibrators that are used for quantitation, e.g., for between-assay normalization, should be run in duplicate. When determining titer from a dilution curve, singlicates can be run if more than two dilution points are used for titer calculations.

Plate Set-up

Bring all reagents to room temperature (18-30° C) equilibration (at least 30 minutes).

- Determine the number of wells for the assay run. Duplicates are recommended, including 8 Calibrator wells and 2 wells for each sample and control to be assayed.
- Remove the appropriate number of microwell strips from the pouch and return unused strips to the pouch. Reseal the pouch and store refrigerated.
- Add 200-300ul Working Wash Solution to each well and let stand for about 5 minutes. Aspirate or dump the liquid and pat dry on a paper towel before sample addition.

Assay Procedure

ALL STEPS ARE PERFORMED AT ROOM TEMPERATURE (25-28° C). After each reagent addition, gently tap the plate to mix the well contents prior to beginning incubation.

1. 1st Incubation [100ul – 60 min; 4 washes]

- Add 100ul of calibrators, samples and controls each to pre-determined wells.
- Tap the plate gently to mix reagents and incubate for 60 minutes.
- Wash wells 4 times and pat dry on fresh paper towels. As an alternative, an automatic plate washer may be used. Improper washes may lead to falsely elevated signals and poor reproducibility.

2. 2nd Incubation [100ul – 30 min; 5 washes]

- Add 100ul of diluted Anti-Rat IgG HRP to each well.
- Incubate for 30 minutes.
- Wash wells 5 times as in step 2.

3. Substrate Incubation [100ul – 15 min]

- Add 100ul TMB Substrate to each well. The liquid in the wells will begin to turn blue.
- Incubate for 15 minutes in the dark, e.g., place in a drawer or closet.

Note: If your microplate reader does not register optical density (OD) above 2.0, incubate for less time, or read OD at 405-410 nm (results are valid).

4. Stop Step [Stop: 100ul]

- Add 100 ul of Stop Solution to each well.
- Tap gently to mix. The enzyme reaction will stop, liquid in the wells will turn yellow.

5. Absorbance Reading

- Use any commercially available microplate reader capable of reading at 450nm wavelength. Use a program suitable for obtaining OD readings, and data calculations if available.
- Read absorbance of the entire plate at 450nm using a single wavelength within 30 minutes after Stop Solution addition. If available, program to subtract OD at 630nm to normalize well background.

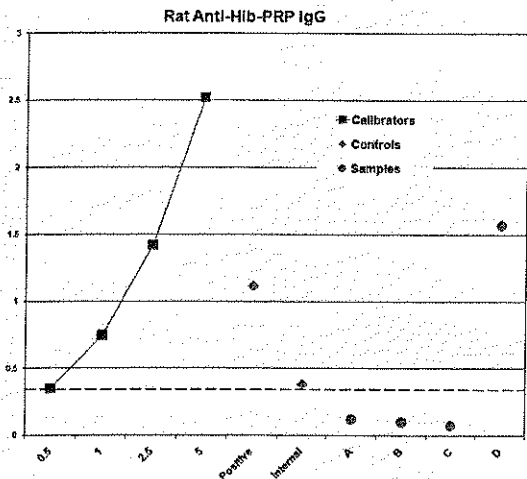
INTERPRETATION OF RESULTS

A. Antibody Activity Threshold Index

Compare Samples to 0.5 U/ml Calibrator or Internal Control

= Positive/Negative Cut-off.

Example:



Results

The **sensitivity** of the assay to detect anti-Hib-PRP IgG, from either natural exposure or vaccination, is controlled so that the **0.5 U/ml Calibrator** represents a threshold OD for most true positives in rat serum diluted to 1:50 or greater. Visual inspection of the data in the above graph shows the following:

Calibrators – dilution curve of antiserum from anti-Hib-PRP immunization, shows the OD range of the assay; high value indicates optimal sensitivity of the assay.

0.5 U/ml: a 'Cut-off' line has been drawn to indicate a threshold distinguishing between **Positive/Negative**. This is not a clear-cut threshold, rather a low OD area that could represent either low positives or high background negatives.

Positive Control – an anti-Hib-PRP serum; net OD > 0.5. This Control can be used to normalize between-assay variation.

Internal Control – a true positive from an immune rat that represents the investigator's experience in distinguishing low positive from negative samples (not in kit). This should be run in each assay to supplement the 0.5 U/ml Calibrator for Positive/Negative discrimination purposes.

Samples A,B,C,D – 3 samples (1:50) (A, B, C) are negative: below the threshold; 1 sample (D) is positive: clearly above the threshold.

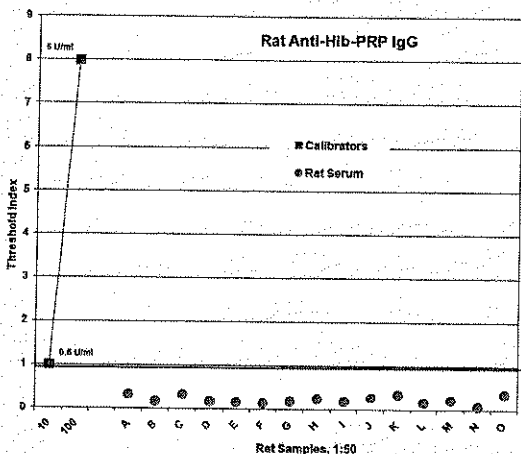
The 0.5 U/ml Calibrator can be used to calculate a **Threshold Index** that numerically discriminates Positive/Negative:

- ❖ Divide each Sample net OD by the 0.5 U/ml Calibrator net OD. Values above 1.0 are a measure of **Positive** Antibody Activity; below 1.0 are **Negative** for antibody.

Example:

Rat Serum IgG

A panel of pooled and individual sera from laboratory rats was tested for anti-Hib-PRP IgG (1:50 dilution). **Threshold Index** was calculated using the 0.5 U/ml Cal.



Results

Anti-Hib-PRP IgG: all sera, pooled [A-D] and individual, were negative (below the 1.0 Threshold Index).

Notes:

1. **Positives** may be due to prior encounter with the virus or non-Hib-PRP proteins with common epitopes..
2. The **sensitivity** of the assay may be adjusted by changing the sample dilutions: a) increase dilution (e.g., 1:100) to lower the signals of borderline positives to negative; b) decrease dilution (e.g., 1:20) to convert borderline samples to positive. With the latter, the values of negatives may increase, so an alternative threshold should be considered using known negatives to develop a **Positive Index** (see below) or use an **Internal Control** (Page 5).
3. When the **Positive Index** is above 5.0, using a dilution curve to calculate titer is a more accurate quantitation method (see Method C).

B. Positive Index

Experimental sample values may be expressed relative to the values of Control or Non-immune samples, by calculation of a **Positive Index**. One typical method is as follows:

1. Calculate the net OD mean + 2 SD of the Control/Non-immune samples = **Positive Index**.
2. Divide each sample net OD by the Positive Index. Values above 1.0 are a measure of **Positive Antibody Activity**; below 1.0 are **Negative** for antibody.

INTERPRETATION OF RESULTS (cont)

A sample value would be **Positive** if significantly above the value of the pre-immune serum sample or a suitably determined non-immune panel or pool of samples, tested at the same sample dilution.

This calculation also **quantifies** the positive Antibody Activity level, assigning a higher value to samples with higher Antibody Activity, and vice versa.

Method C. Titers from Sample Dilution Curves

The titer of elevated antibody activity calculated from a dilution curve of each sample is recommended as the most accurate quantitative method. Best precision can be obtained using the following guidelines:

1. Use an OD value Index in the mid-range of the assay (2.0 – 0.5 OD); this provides the best sensitivity and reproducibility for comparing experimental groups and replicates. An arbitrary 1.0 OD is commonly used.
2. Prepare serial dilutions of each sample to provide a series that will produce signals higher and lower than the selected index. With accurate diluting, duplicates may not be required if at least 4 dilutions are run per sample.
3. A 5-fold dilution scheme is useful to efficiently cover a wide range which produces ODs both above and below 1.0 OD. The dilution scheme can be tightened to 3-fold or 2-fold for more precise comparative data.
4. The Positive and Sensitivity Control values can be used to normalize inter-assay values.

Calculations

1. On a log scale of inverse of Sample Dilution as the x-axis, plot the OD values of the two dilutions of each positive sample having ODs above and below the OD value of the Index (arbitrary or selected Calibrator).
2. From a point-to-point line drawn between the two sample ODs, read the dilution value (x-axis) corresponding to the OD of the selected Index

= IgG Antibody Activity Units

Assay Sensitivity

The HbO-HA coating level, HRP conjugate concentration and Low NSB Sample Diluent are optimized to differentiate anti-Hib-PRP IgG from background (non-antibody) signal with rat serum samples diluted 1:50.

PRECAUTIONS AND SAFETY INSTRUCTIONS

Calibrators, Sample Diluent, and Antibody HRP contain bromonitrodioxane (BND: 0.05%, w/v). Stop Solution contains dilute sulfuric acid. Follow good laboratory practices, and avoid ingestion or contact of any reagent with skin, eyes or mucous membranes. All reagents may be disposed of down a drain with copious amounts of water. MSDS for TMB, sulfuric acid and BND can be requested