

## *n*Taq

Cat.#	Size	Conc.
P025A	250 units	5 units/ $\mu$ l
P050A	500 units	5 units/ $\mu$ l

Store at -20°C

Supplied with: 10X *n*Taq Buffer (Mg<sup>2+</sup> plus)  
dNTP Mixture  
Sterile water

### India Contact:

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### Product description

A gene encoding the *Thermus aquaticus* (Taq) DNA polymerase was cloned and expressed in *E. coli*, and the enzyme was purified to homogeneity. The purified Taq polymerase (*n*Taq) has optimal activity at high temperatures (72°C), which helps amplify secondary-structured regions. *n*Taq DNA Polymerase has greatly reduced activity of 5'→3' exonuclease activity. As a result, formation of erroneously amplified products caused by the 5'→3' exonuclease activity of wild type polymerase is effectively prevented. In addition, high GC-content or secondary-structured regions can be efficiently amplified due to the genetic modifications in the catalytic domain of the polymerase.

### Characteristics

- Molecular weight: 94 kDa
- Error rate: 2.4 X 10<sup>-5</sup>
- Thermal stability: Half life of 40 min at 95°C
- A-tail formation at 3' ends of amplified duplex DNA.

### Applications

- Amplification of DNA fragments shorter than 3 kb (Suitable for general PCR analysis)
- Amplification of cDNA and genomic DNA.
- Primer extension
- Colony PCR

For Research Use Only. Not for use in diagnostic procedures.

ISO9001 ISO14001 ISO13485

- Multiplex PCR
- Labeling of DNA fragments with radioactive-isotopes
- Nucleotide sequencing

### Quality control

- Purity: >99% on SDS-PAGE
- Endonuclease-free
- Exonuclease-free
- RNase-free
- Inhibitor-free

### Unit definition

One unit is defined as the amount of enzyme required to incorporate 10 nmol of dNTP into acid insoluble materials in 30 min at 74°C in a 50- $\mu$ l reaction mixture (20 mM Tris-HCl/pH 8.8, 50 mM KCl, 2.5 mM MgCl<sub>2</sub>, 10 mM  $\beta$ -mercaptoethanol, 12.5  $\mu$ g of calf thymus DNA).

### Storage buffer

20 mM Tris-HCl (pH 7.9), 100 mM KCl, 0.1 mM EDTA, 1 mM DTT, 0.5% NP-40, 0.5% Tween-20, 50% glycerol.

### 10X *n*Taq Buffer (Mg<sup>2+</sup> plus)

Mg<sup>2+</sup> free Buffer: containing 15 mM MgCl<sub>2</sub>  
Mg<sup>2+</sup> plus: MgCl<sub>2</sub> free

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Mg<sup>2+</sup> plus: MgCl<sub>2</sub> free

### Standard PCR conditions

- PCR mixture<sup>a</sup>

10X <i>n</i> Taq Buffer (Mg <sup>2+</sup> plus)	2 $\mu$ l
<i>n</i> Taq DNA Polymerase <sup>b</sup> (5 units/ $\mu$ l)	0.2 $\mu$ l
dNTP Mixture (2 mM each)	2 $\mu$ l
Template DNA <sup>c</sup> (0.1~500 ng/ $\mu$ l)	1 $\mu$ l
Primer 1 (5 pmole/ $\mu$ l)	1 $\mu$ l
Primer 2 (5 pmole/ $\mu$ l)	1 $\mu$ l
Sterile water	up to 20 $\mu$ l
<sup>a</sup> Assemble the reaction mixture on ice	
<sup>b</sup> Add the PCR polymerase at the final step	
<sup>c</sup> Plasmid DNA, 0.1 ng~30 ng; genomic DNA, 50 ng~500 ng	

- PCR cycle

Initial denaturation	95°C	2 min
Denaturation	95°C	30 sec
Annealing <sup>a</sup>	55°C~65°C	30~60 sec
Elongation	72°C	1 min/kb
Number of cycles	25~35 times	
Final elongation	72°C	5 min
When cycles are over, keep the reaction mixture at 4°C; may add 10 mM EDTA until use to prevent DNA degradation.		
<sup>a</sup> Recommended annealing temperatures is 5 to 10°C below the lower T <sub>m</sub> of the two primers used.		

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