

## AmplifyRP® Acceler8® for Citrus Greening (Las)

### Rapid DNA Test Kit

Product No. ACS 15400/0008



### Intended Use:

AmplifyRP Acceler8 for Las is a rapid DNA amplification and detection platform designed for field-based or laboratory testing of citrus crops for *Candidatus Liberibacter asiaticus*, the causal agent of HLB (Citrus Greening).

This assay does not cross-react with *Candidatus Liberibacter americanus*, *Candidatus Liberibacter africanus*, *Candidatus Liberibacter solanacearum*, or *Xanthomonas citri*.

### Kit Storage:

All kit components should be stored refrigerated (2 - 8 °C).

Before use, allow all kit components to warm to room temperature (18 - 30 °C) for 20 to 30 minutes.

### Sample Preparation

*NOTE: AmplifyRP is a very sensitive molecular assay. Do not re-use disposable kit components. It is recommended that latex gloves be worn when taking samples and performing assay. If wearing latex gloves, change them between samples and test runs. Sanitize work area and non-disposable equipment between runs with a 10 % bleach solution.*

1. Symptoms of HLB in an infected plant may include, but not limited to, blotchy leaf mottling, vein yellowing, misshapen fruit, and / or fruit drop. HLB is a phloem limited pathogen making it important to select the right sample for testing. For best results, Agdia recommends testing the mid-rib of suspect leaf samples.
2. Cut a section of mid-rib (Figure 1) from a symptomatic leaf that is approximately 0.06 g in weight. Make certain you clean cutting instruments between samples with a 10 % bleach solution to avoid contamination.
3. Insert the sample between the mesh linings of the sample extraction bag. Add 10 drops\* (~300 µL) of AMP1 extraction buffer to the tissue inside the bag. The sample can be extracted by rubbing the outside of the bag with a blunt object such as a pen or marker on a hard surface (Figure 2). Once the sample has been thoroughly homogenized, it is ready to be tested.

\*NOTE: This test was optimized using a 1:5 tissue to buffer ratio for sample extraction.

### Amplification

1. Allow heat block to warm to 39 °C before preparing reactions. If using an Agdia supplied heat block, allow 2 to 3 minutes for this step.
2. Remove the strip of reaction pellets from the desiccated container included in the kit. While securing the strip of pellets in a 200 µL PCR tube rack, cut the number of reaction pellets from the strip that are intended for use. Immediately place remaining reaction pellets back into the desiccated tube for later use.
3. Dispense 10 µL of PD1 Pellet Diluent into a reaction pellet. Be sure the pellet is visible in the bottom of the tube before opening or dispensing liquid into the tube.
4. Using a 1 µL loop or pipette, immediately transfer 1 µL of sample extract into the rehydrated reaction pellet. Recap the tube and mix by flicking the bottom of the tube 6 to 8 times or by using a laboratory vortex mixer. Then shake or centrifuge the liquid contents to the bottom of the tube.
5. Add reaction to the portable heat block for 20 minutes.
6. Immediately remove reaction from heat block and proceed to detection steps.

### Contents of Kit:

- Reaction pellets (8)
- Amplicon detection chambers (8)
- PD1 pellet diluent (1.0 mL)
- AMP1 extraction buffer (20 mL)
- Sample extraction bags, mesh (8)
- 1 µL transfer loops (10)

### Not Included but Required:

- Portable heat block (ACC 00592)
- 10 µL pipette (ACC 00770/0010)
- 10 µL pipette tips (ACC 00160)
- 200 µL PCR tube rack (ACC 00170)

\* A starter pack inclusive of the items not included above can be purchased from Agdia (ACC 00150).

Figure 1. HLB Infected Leaf



Figure 2. Sample Extraction



## Detection

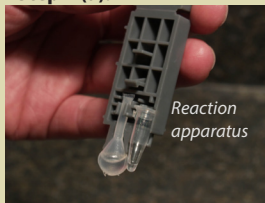
**In order to avoid possible contamination of future tests, DO NOT open the reaction pellet.**

1. Open the foil pouch containing the Amplicon detection chamber. There are two pieces to the chamber as indicated in the figure to the right.
2. a.) Add the unopened reaction tube to reaction apparatus as illustrated to the right. b.) Once the tube has been added, snap the apparatus shut which will immobilize the reaction tube.
3. Add the reaction apparatus to the detection chamber housing as indicated. **IMPORTANT: The reaction tube should be facing toward the lateral flow strip, contained in the housing, during this step.**
4. Push down on the handle of the detection chamber housing until it snaps shut. Wait 20 minutes before interpreting results. Positive results may be visible in as little as 5 to 10 minutes. Samples that contain lower copy numbers may take up to 20 minutes to produce a positive test line.

Step 1.



Step 2 (a).



Step 2 (b).



Step 3.



Step 4.

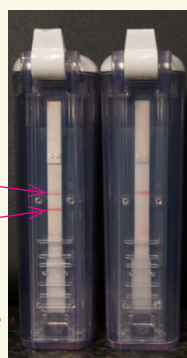


## Interpret Results

Result	ImmunoStrip Reaction
<b>Positive</b>	Control and Test lines are both visible.
<b>Negative</b>	Control line is visible. Test line not visible.
<b>Invalid</b>	Control line not visible.

Control Line  
Test Line

Positive  
(+)



Negative  
(-)

## Limitations

The following is a description of factors that could limit test performance or interfere with proper test results.

**Sample Extraction Buffer:** This test must be used with the supplied sample extraction buffer to obtain optimal results.

**Addition of sample extract to reaction pellet:** It is important to add only the prescribed amount of sample extract to reaction pellets. Adding too much extract may cause test failure.

**Storage:** Test results may be weak or the test may fail if the storage instructions are not followed properly. The lyophilized test components must remain protected from light to prevent bleaching and sealed with desiccant when not in use to prevent moisture degradation, which may affect test results. Do not store pellets at temperatures greater than 42 °C, as this may cause test failure.

## Questions or Technical Support:

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